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CONIDIA OF ACROSPERMUM COMPRESSUM AND A. GRAMINUM

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(With 2 Text-figures)

Acrospermum compressum and A. graminum, which have earlier been considered as synonyms, produce different types of conidial apparatus in culture. A. compressum forms two-celled conidia, whilst A. graminum forms unicellular conidia.

Acrospermum compressum Tode ex Fr. and A. graminum Lib. show characteristic differences when grown in culture. The two species have, however, been regarded as one. Rehm (1896), Riddle (1920) and Brandriff (1936) regarded A. graminum as a variety of A. compressum. This view is based on the similarity of fruit-bodies of the two species, but no cultural evidence was put forward to support it, although Brandriff grew A. compressum. She states that 'after three weeks fruiting structures developed. These had the appearance of spermogonia or very small pycnidia. They were provided with short beaks, and contained minute unicellular bodies of the aspect of spermatia.' No drawings or measurements of these structures were given. Cultures from recent British collections differ from those described by Brandriff.

ACROSPERMUM COMPRESSUM

Isolations of A. compressum have been made from the following five collections:

Herb. University Sheffield number	Host	Locality	Date
1178	Urtica dioica	Howldale, Pickering, Yorks, Coll. W. G. Bramley	13. vi. 54
1643 <i>a</i>	U. dioica	Middle marsh, Wheatfen Broad, Surlingham, Norfolk	12. iv. 55
1647	U. dioica	Lowdales, Hackness, Yorks	16. iv. 55
1648	Galium aparine	Lowdales, Hackness, Yorks	16. iv. 55
1649	Urtica dioica	Forge Valley, Yorks.	15. iv. 55

Isolations from all these collections produced a similar type of growth in culture and the description below is based on specimen no. 1643*a*.

Fruit-bodies (Fig. 1 Å): single or in groups, club-shaped to cylindrical, slightly compressed, pointed, pale yellow to brown and somewhat translucent when moist, darker on drying, usually 2-3 mm. high and $200-400 \mu$ broad. The wall of the fruit-body is about $50-60 \mu$ thick, composed of parallel narrow hyphae, turning outwards and expanding at their tips to form a pseudoparenchymatous outer layer of brown polygonal cells



Fig. 1. Acrospermum compressum. A, B, C from Herb. University Sheffield, no. 1643*a*; D from 1178. A, fruit-bodies on Urtica stem; B, asci and ascospores; C, conidia from a single ascospore culture, 14 days old on maize-extract agar; D, conidia from a similar culture derived from Herb. no. 1178.

about 10 μ wide. Asci (Fig. 1B): confined to the wider upper part of the fruit-body, fasciculate, narrowly cylindrical, up to 500 μ long, $3-4\mu$ wide, separated by numerous filiform paraphyses. Ascospores: 8, filiform, hyaline, almost as long as the asci, about 0.5 μ wide.

This specimen has been matched with Schmidt and Kunze Exsiccatum no. 69, also on *Urtica dioica*, which was one of the exsiccati cited by Fries (1822).

Cultures. From a suspension of crushed asci, single ascospore and single ascus cultures were prepared. On maize-extract agar the fungus grew slowly, forming flat whitish colonies. The hyphae were at first confined to the medium, $1-3\mu$ wide, often forming characteristic coiled skeins. Later aerial hyphae were produced, and these bore conidiophores within 10-14 days of inoculation. The conidiophores were mostly unbranched and consisted of pale yellow upright hyphae 20-80 μ long, 3-4 μ wide at the base, tapering to 2μ near the apex. The tip of the conidiophore bore a radiate group of cylindrical conidia, and up to fifteen conidia have been counted on a single head (Fig. 1 C, D). The first conidium arises as the blown-out tip of the conidiophore. Subsequent conidia are formed close to it (probably acrogenously) so that a close crown of conidia arises. When a conidium is detached an inconspicuous flattened scar is visible at the conidiophore apex. Mature conidia are pale yellow in mass, cylindrical to club-shaped, tapering to a flattened scar below, rounded above, usually with a single median transverse septum, slightly constricted. Occasional 2- to 3-septate spores were found, and in old cultures the individual cells of the spores were often inflated. The ranges in size of the conidia from the five collections were: 1178, 10–20 \times 3–4 μ ; 1643 a, $13-23 \times 2 \cdot 5 - 3 \mu$; 1647, 12-30 × 3-4 μ ; 1648, 12-18 × 2 \cdot 5 - 3 μ ; 1649. $8-18 \times 3-4 \mu$.

Acrospermum graminum

Isolations of *Acrospermum graminum* have been made from the following four collections:

Herb. University Sheffield Date Host Locality number Bolton Percy, Yorks. Coll. W. G. Bramley 365 Wheat stems 1. v. 50 Fulwood, near Sheffield Deschampsia caespitosa 5. vi. 51 1055 Kilnsea, E. Yorks 21. vii. 49 Agropyron repens 799 Home Marsh, Wheatfen Broad, Surlingham, 1644 Phragmites communis 12. iv. 55 Norfolk

All isolates produced similar growth in culture. The following description is based on specimen no. 365.

Fruit-bodies (Fig. 2A): club-shaped, $1-1\cdot 2$ mm. high and 200-300 μ wide, yellow to brown in colour, single or in groups of two to four, seated on a broader base and bursting through the epidermis of the leaf sheath, rounded or pointed at the apex, slightly flattened above, collapsing on drying to form shallow furrows. The wall of the fruit-body is made up of narrow parallel, interwoven hyphae, turning outwards to form rounded or polygonal yellowish brown cells about 6μ wide. Asci: fasciculate,

narrowly cylindrical, tapering at the base and rounded above, $400-600 \mu$ long, $4-6 \mu$ wide, separated by filiform paraphyses. Ascospores: 8, filiform, hyaline, about as long as the asci, 0.5μ wide.

The specimen has been matched with Libert's Pl. Crypt. Arduen. no. 33.

Cultures. Six cultures were prepared from ascospores and asci. The asci and ascospores germinated readily and were transferred to a variety



Fig. 2. Acrospermum graminum. Herb. University Sheffield no. 365. A, fruit-bodies on wheat stems; B, asci and ascospores; C, conidia from single ascospore culture, 14 days old on maize-extract agar.

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of agar media. The fungus grew rather slowly, producing regular circular colonies with a submerged cream-coloured mycelium, and an aerial growth which was faintly green in colour, forming rope-like strands. Within 10 days conidia were produced on short conidiophores projecting from the rope-like strands of aerial mycelium or from the submerged mycelium. Conidia were produced in all cultures, including one derived from a single ascospore and one from a single ascus. The conidiophores are tapering and unbranched and occasionally bear a septum at the base. are $15-16 \mu \log \times 2.5-4 \mu$ wide at the base tapering to $1.5-2 \mu$ at the apex. The first formed conidium is produced terminally as a cylindrical swelling at the tip of the conidiophore and is displaced laterally by the formation of a second growing point, which grows beyond the first formed conidium and produces a second conidium. This process is repeated so that an 'ear' of conidia is formed. Eventually in culture, hundreds of conidia may be formed in a single head. When conidia are displaced a series of small peg-like scars can be observed on the conidiophore (Fig. 2C). The conidia are hyaline, cylindrical or elliptical, and measure $3-6 \times 1.5-2.5 \mu$ (mostly $4 \times 1.5 \mu$). Similar conidiophores and conidia have also been found developing from the wall of the fruit-body, after incubation in a damp-chamber.

No appreciable difference has been detected in the growth and dimensions of the other isolates. In addition to these isolates, the fungus has been isolated repeatedly from the washed fragments of upper internodes of standing stems of *Agropyron repens*, collected at Lindrick Common, near Worksop by Mr H. J. Hudson and from fragments of standing stems of *Dactylis glomerata*.

Zogg (1950) records isolating Acrospermum compressum var. graminum from cereals, but does not describe his isolates.

DISCUSSION

Comparison of over fifty collections of Acrospermum on herbaceous stems and grasses from Herb. Kew. showed great variation in size and shape. However, the height of fruit-bodies in collections on grasses rarely exceeds 1.5 mm., whilst on herbaceous stems they are frequently larger, often up to 3 mm. The similarity of fruit-bodies on the two types of host has doubtless led earlier investigators to regard them as representing a single species. However, the differences in conidial apparatus suggest that they should be regarded as separate species. It would be of interest to study isolates of Acrospermum on hosts such as Typha and Carex, on which it is known to occur, to see which species is present. A. foliicolum Berk., which grows on leaves of deciduous trees in the Southern states of North America, resembles A. compressum, and Riddle (1920) has regarded it as a variety of the latter. It would be of interest to study isolates of this fungus.

The discrepancy between Brandriff's description of conidia of A. compressum and the present isolates is puzzling. In none of the British isolates has pycnidia been observed, although the fungus has been kept in culture for several months. I have examined Brandriff's collections and slides. Her collections on *Caltha* are more robust than those on grasses and sedges, and it seems to me probable that the former should be ascribed to A. compressum and the latter to A. graminum.

The taxonomic position of Acrospermum has puzzled most authorities. Rehm (1896) has reviewed earlier dispositions of the genus. Saccardo (1883) classified it with the Hysteriaceae-Scolecosporae, dividing the group into two series: (A) Genuinae and (B) Desciscentes (lit. degenerate, spurious), placing Acrospermum in the latter group. Rehm himself erected the group Pseudohysterineae for the genus. Brandriff (1936) also indicated relationship to the Hysteriales. Luttrell (1955) places the genus in the Coryneliales, but suggests that it should possibly be considered 'as a partially reduced perithecial form and placed in the vicinity of the Clavicipitaceae'. However, the asci of Acrospermum do not possess the characteristic apical apparatus found in Claviceps, and a satisfactory disposition of the genus is still awaited.

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